Ageratum conyzoides L., belonging to the family Asteraceae, is an annual herbaceous plant and is an invasive weed. It contains bioactive compounds that are reported to possess therapeutic properties. The species has been studied widely owing to its biological properties and its potential application in medicine and agriculture. It is used in the treatment of burns and wounds, arthrosis, malaria, asthma, leprosy and dermatitis. It also has insecticidal activity against a range of major pests of field crops. Chromatographic analyses have identified pyrrolizidine alkaloids, phenolic acids, coumarin and polymethoxyflavones. The present study was conducted to determine the qualitative and quantitative properties of the crude extracts of the leaves. Extracts were prepared by using petroleum ether, chloroform and methanol solvents. The yield of extract was calculated for all the three solvents and they are studied for qualitative analysis of phytochemical compounds and quantitative analysis. The qualitative phytochemical tests exhibited the presence of alkaloids, carbohydrates, phenols, tannins, steroids, terpenoids, glycosides, saponins, and amino acid. Quantitative analysis suggests the presence of tannins, alkaloids and phenols in the leaves and this can be utilized for further investigations.

Keywords: Ageratum conyzoides L., phytochemical, bioactive, quantitative, tannins.

INTRODUCTION

Since ancient times, medicinal plants have been used for their healing properties and they remain the fundamental sources of novel biological active compounds (Bearth et al., 2014). Medicinal plants and natural products possessing pharmacological models have been considered as alternative therapy for the treatment of various diseases (Singh et al., 2017). The chemical compounds produced by plants are as a result of normal metabolic activities. These chemical compounds are classified into primary and secondary metabolites, and the secondary metabolites and other chemical constituents contribute to their medicinal value (Varadarajan et al., 2008).

Phytochemicals are non-nutritive plant chemicals that have protective and preventive properties against diseases (Breslin and Andrew, 2017). They are among the secondary metabolites naturally occurring in plants and most often possessing health benefits (Kabera et al., 2014). They play a vital role in the protection of plants as antibacterial, antiviral, antifungal and insecticidal agents (Hajlaoui et al., 2009).

Ageratum conyzoides L. belongs to the family Asteraceae. It is native to Central America and is invasive in Southeast Asia, West Africa, South China and India (Iwu, 2000; Prince and Prabakaram, 2011; Amadi et al., 2012). It is an annual herbaceous plant which grows to approximately one meter in height. The stems and leaves are hairy, the leaves are ovate bearing purple to white flowers (Marks and Nwachuku, 1986). The plant grows widely in agricultural land, roadside and is very common in waste places and on ruined sites. It is commonly utilized as folk remedies for the treatment of pneumonia, cure wounds and burns (Durodola, 1977), colic, colds, and fevers, diarrhoea, rheumatism, spasms (Oliveira et al., 1993), have anti-inflammatory, antipyretic, analgesic
activity (Abena et al.), insecticidal activity (Gbolade et al., 1999), antidiysenteric, and antilithic activity (Borthakur and Baruah, 1987). The plant extract is found to possess cardiovascular depressant (Achola et al., 1994) and antioxidant activities (Amal et al., 2010).

The present study investigated the qualitative and quantitative analysis of phytochemicals of crude extracts of A. conyzoides leaves.

**MATERIALS AND METHODS**

**Plant material**

Fresh plants of *Ageratum conyzoides* L. was collected from Zemabawk, Aizawl, Mizoram, India and authenticated by a taxonomist at the Botanical Survey of India, Shillong, India, vide letter No.BSI/ERC/Tech./Plan Iden./2018/136. A voucher specimen has been deposited in the Department of Pharmacy, RIPANS. Fresh leaves were washed with tap water and then finally with distilled water followed by shade drying at room temperature for 15-20 days. The shade-dried leaves were made into powdered using electrical blender.

**Preparation of extract**

The dried leaves powdered were introduced to successive extraction by refluxing in the Soxhlet apparatus using petroleum ether, chloroform and methanol as solvents. About 245 g of powdered leaves were uniformly packed into the thimble and extracted with the above solvents. The process of extraction continues for 36 hours or till the solvent in the siphon tube become colorless. The extract was taken into sterile beaker and concentrated by evaporating the solvents in water bath. The dried extract was stored in refrigerator for further analysis.

**Qualitative phytochemical analysis**

The crude methanolic extract of *Ageratum conyzoides* L. leaves were screened for phytochemicals such as alkaloids (Dragendorff’s test, Mayer’s test, Wagner’s test), carbohydrates (Fehling’s test, Benedict’s test), phenols and tannins (ferric chloride test, Lead acetate test), flavonoids (Shinoda test, lead acetate test, alkali test), saponins (foam test), triterpenoid (Salkowski’s test), glycosides (Liebermann’s test, Keller-Kilani test ), steroids, proteins (Millon’s test, ninhydrin test) (Aziz, 2015; Yadav and Agarwala, 2011; Thakur, 2018; Santhi and Sengottuvel, 2016).

**Quantification of alkaloids**

1 mg of the crude methanolic extract of *A. conyzoides* leaves was dissolved in dimethyl sulphoxide (DMSO), 1 ml of 2N HCl was added and allow to filtered. 5 ml each of bromocresol green and phosphate buffer were added to the filtrate and transferred to separating funnel. 1, 2, 3 and 4 ml of chloroform was added to solution mixture by vigorous shaking and collected in 10 ml volumetric flask. Then, the volume of these flasks was diluted to the mark with chloroform. A set of reference standard solutions of atropine (20, 40, 60, 80 and 100 µg/ml) were prepared in the similar manner as described above. Absorbance for test and standard solutions were determined against the blank at 470 nm using UV-visible spectrophotometer. The total alkaloid content was expressed as mg of AE/gm of extract (Shamsa et al., 2008; Raob et al., 2016).

**Quantification of phenols**

Folin-Ciocalteu spectrophotometric method was used to determined total phenolic content in crude methanolic extract of *A. conyzoides* leaves. 1 ml of extract and 9 ml of distilled water was taken in 25 ml of volumetric flask, followed by addition of 1 ml of Folin-Ciocalteu phenol reagent and shaken well. After a period of 5 minutes, 10 ml of 7% sodium carbonate solution was added to the mixture. The volume was made up to 25 ml with distilled water and the absorbance was determined at 765 nm. A standard curve was plotted using gallic acid.

**Figure 1:** Standard curve for total alkaloidal content.

**Figure 2:** Standard curve for total tannin content.
water. A set of reference standard solutions of gallic acid (20, 40, 60, 80 and 100 µg/ml) were prepared in the same manner as described above. The prepared test and standard solutions were incubated for 90 minutes at room temperature. The absorbance for test and standard solutions were recorded against blank at 550 nm. The total phenolic content was expressed as mg of GAE/gm of extract (Rasool et al., 2011; Ghasemzadeh et al., 2010; Stankovic, 2011).

Quantification of tannins

Total tannin content was determined by Folin-Ciocalteu spectrophotometric method using gallic acid as standard. About 0.1 ml of plant extract and 7.5 ml of distilled water were taken in 10 ml volumetric flask, 0.5 ml of Folin-Ciocalteu phenol reagent and 1 ml of 35% Sodium carbonate solution was added. The volume was made up to 10ml with distilled water. The solution mixture was shaken well and kept aside for 30 minutes at room temperature. A set of standard solutions of gallic acid (20, 40, 60, 80 and 100 µg/ml) were prepared in the similar manner as mentioned above. The absorbance for test and standard solutions were measured against blank at 725 nm. The total tannin content was expressed as mg of GAE/gm of extract (Marinova et al., 2005; Singh et al., 2012; Afify et al., 2012; Miean and Mohamed, 2011).

RESULTS

Extraction yield

245 gm of dried weight of A. conyzoides leaves were utilized for extraction using different solvent such as petroleum ether, chloroform and methanol and the extractive yield was summarized in Table 1.

Table 1: Extraction yield of the leaves of Ageratum conyzoides L.

<table>
<thead>
<tr>
<th>S.No</th>
<th>Extract</th>
<th>Extractive yield (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Petroleum ether</td>
<td>5.52</td>
</tr>
<tr>
<td>2</td>
<td>Chloroform</td>
<td>6.55</td>
</tr>
<tr>
<td>3</td>
<td>Methanol</td>
<td>8.39</td>
</tr>
</tbody>
</table>

Qualitative phytochemical analysis

The qualitative analysis of phytochemicals from crude methanolic extract of A. conyzoides leaves was analyzed and showed the presence of several phytoconstituents which are summarized in Table 2. The results revealed the presence of alkaloids, carbohydrates, phenols and tannins, flavonoids, saponins, triterpenoids, glycosides and steroids.

Table 2: Phytochemicals in the crude of methanolic extract of Ageratum conyzoides L. (+ Present; - Not detected)

<table>
<thead>
<tr>
<th>S.No</th>
<th>Phytochemicals</th>
<th>Methanolic extract</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Alkaloids</td>
<td>+</td>
</tr>
<tr>
<td>2</td>
<td>Carbohydrates</td>
<td>+</td>
</tr>
<tr>
<td>3</td>
<td>Phenols and tannins</td>
<td>+</td>
</tr>
<tr>
<td>4</td>
<td>Flavonoids</td>
<td>+</td>
</tr>
<tr>
<td>5</td>
<td>Saponin</td>
<td>+</td>
</tr>
<tr>
<td>6</td>
<td>Triterpenoids</td>
<td>+</td>
</tr>
<tr>
<td>7</td>
<td>Glycosides</td>
<td>+</td>
</tr>
<tr>
<td>8</td>
<td>Steroids</td>
<td>+</td>
</tr>
<tr>
<td>9</td>
<td>Proteins</td>
<td>-</td>
</tr>
</tbody>
</table>

Quantification of phytochemicals

The quantitative estimation of crude methanolic extract of A. conyzoides leaves was examined for total alkaloids, total tannins and total phenolic content as mentioned in Table 3. The alkaloid content in plant extract was expressed as atropine equivalent (mg of AE/g of extract) and was found to be 8.7±0.2 mg of AE/g of extract. Total phenolic and tannin were estimated using Folin-Ciocalteu reagent and were expressed as gallic acid equivalent (mg of GAE/g of extract). The total tannin con-
tent was estimated to be 108.17±0.76 mg of GAE/g of extract, while that of total phenolic content was examined to be 122.08±1.91 mg of GAE/g of the extract.

**DISCUSSION**

When herbs are used for treating certain ailments and disorder, phytochemicals as bioactive compounds are responsible for its medicinal as well as physiological activities (Alabri et al., 2014). Qualitative phytochemical analysis of the present study revealed the presence of alkaloids, carbohydrates, phenols and tannin, flavonoids, saponins, triterpenoids, glycosides and steroids. Alkaloids possess biological properties such as analgesic, antibacterial, antispasmodic activities.

Phenolic compounds show varieties of pharmacological activities such as antiapoptosis, antiaging, anticarcinogen, antiinflammation, antiatherosclerosis and tannins have astringent property. Flavonoids are commonly used natural antioxidant having antimicrobial as well as anticarcinogenic properties. Saponins are characterized by formation of foam with aqueous solution and are known to possess anti-inflammatory activity (Yadav and Agarwala, 2011). Glycosides have the property of cardiovascular disease (Brain et al., 1985). Quantitative estimation of total alkaloid, tannin and phenol revealed that there is significant presence of tannin and phenol in the plant extract while alkaloid are present in lesser amount.

**CONCLUSION**

Thus from the present study, the crude methanolic extract of *A. conyzoides* leaves possess different varieties of phytochemicals such as alkaloids, carbohydrates, phenols and tannin, flavonoids, saponins, triterpenoids, glycosides and steroids. The study revealed the basis of its use as folk medicine against various diseases due to the presence of these phytochemicals as bioactive compound. So, the plant extract can be utilized for further investigations.

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