

# Bioinformatics Analysis of the Gene *CYP83B1* in Cabbage (*Brassica oleracea* var. *capitata*)

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**Abstract:** CYP83B1 is an important glucosyltransferase enzyme in glucosinolate biosynthesis. Here, the *Brassica oleracea* var. *capitata* *CYP83B1* (*BocCYP83B1*) gene sequence was obtained from *Brassica* database (BRAD), and preformed for bioinformatics analysis. The *BocCYP83B1* gene mapped to chromosomes 8, and contains an open reading frame of 1,473 bp that encodes a 490-amino acid protein with a calculated molecular mass of 55.89 kD and an isoelectric point (pI) of 8.89. Subcellular localization predicted the *BocCYP83B1* gene was in the chloroplast. The conserved domain of the *BocCYP83B1* protein is belonged the p450 superfamily. The CYP83B1 protein is most closely related to *B. rapa*. The findings of the present study provide a molecular basis for the elucidation of *CYP83B1* gene function in cabbage.

## 1. Introduction

Cabbage (*Brassica oleracea* var. *capitata*) is a member of the Brassicaceae family that is widely distributed in the world. In China, cabbage is an important vegetable crop, and consumed considerable every year. Cabbage is generally grown for its leafy head as common edible part, which are crispy, tender, and tasty [1]. Besides its good flavor, cabbage is also a rich source of nutrients, antioxidants, and anticarcinogenic compounds, including carbohydrates, vitamin C, carotenoids, and glucosinolates [1-2].

Glucosinolates are a group of sulfur- and nitrogen-containing secondary metabolites that are mainly found in the order of Brassicales and related groups of dicotyledonous angiosperms [3-4]. Glucosinolates and the hydrolytic myrosinase ( $\beta$ -thioglucoside glucohydrolase) are stored separately under normal situations, but they come into contact with each other when tissues are damaged, and then the glucosinolates are hydrolyzed into several degradation products, such as isothiocyanates and nitriles [5]. Glucosinolates and their degradation products have diverse biological functions, which contribute to human health, as well as the taste and odor of cruciferous crops. The anticancer activity of isothiocyanates has been widely studied, and the mechanism involved has been elucidated [6].

Glucosinolate metabolism in plants is modulated by numerous biotic and abiotic factors, and the regulatory network of glucosinolate metabolism has been well elucidated in *Arabidopsis* [6]. CYP83B1 is metabolized for aromatic oximes, and has higher affinity, particularly in the case of indole-3-acetaldoxime [7]. The gene encoding the CYP83B1 protein has been isolated in *Arabidopsis thaliana* and Chinese cabbage [7]. To date, research studies on *CYP83B1* in cabbage are limited. In the present study, the *CYP83B1* gene sequence of cabbage was obtained from web database, and then bioinformatics analysis of the *CYP83B1* gene were analyzed. The present study aimed to establish the foundation for further studies on the molecular mechanism of *CYP83B1* in cabbage.

## 2. Materials and methods

### 2.1 Sequence Obtain of the *BocCYP83B1* Gene.

The genomic DNA and mRNA sequences of *CYP83B1* gene of cabbage were downloaded and obtained from The *Brassica* database (BRAD) (<http://brassicadb.org>), and then used to subsequent bioinformatic analysis.

### 2.2 Bioinformatics Analysis of the *BocCYP83B1* Gene.

The amino acid sequence, protein molecular weight, isoelectric point, stability index, and hydrophobicity of the *BocCYP83B1* gene were analyzed and predicted by ExPASy (<http://web.expasy.org>) and NCBI (<https://www.ncbi.nlm.nih.gov/>). Subcellular localization was predicted by WoLF PSORT (<http://www.genscript.com/wolf-psort.html>). The conserved domain were predicted by NCBI (<https://www.ncbi.nlm.nih.gov/Structure/cdd/wrpsb.cgi>). Phylogenetic tree analysis of the CYP83B1 proteins was executed in MEGA 6.0 using the neighbor-joining (NJ) method.

## 3. Results

### 3.1 Analysis on Genomic Organization.

The *Brassica* database (BRAD) was used to analyze the chromosomal localization and genomic organization of *BocCYP83B1*. The gene ID in BRAD is Bol033477. The *BocCYP83B1* gene was mapped to chromosomes 8 and has 2 exons and 1 intron (Fig. 1).

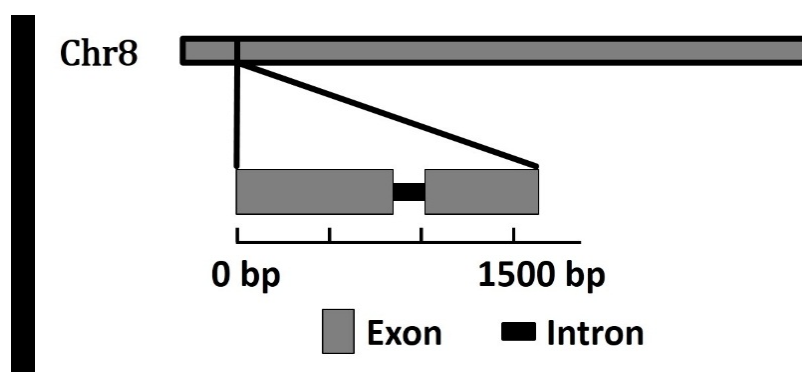


Fig. 1 Chromosomal location and genomic structure of *BocCYP83B1*.

### 3.2 Protein Physical and Chemical Properties Analysis.

Sequence analysis indicated that the *BocCYP83B1* gene contained a 1,473-bp open reading frame (ORF), which encoded a 490-amino acids protein with a calculated molecular mass of 55.89 kD and an isoelectric point (pI) of 8.89. The amino acid types and proportions of the *BocCYP83B1* gene was shown in Figure 2, the highest number of amino acid is Leucine (Leu), whereas the lowest number is Tryptophan (Trp). Its predicted formula was  $C_{2538}H_{3994}N_{664}O_{703}S_{26}$ . Its total average hydrophilicity index was -0.158, liposoluble index was 89.78, and instability index in solution was 36.88.

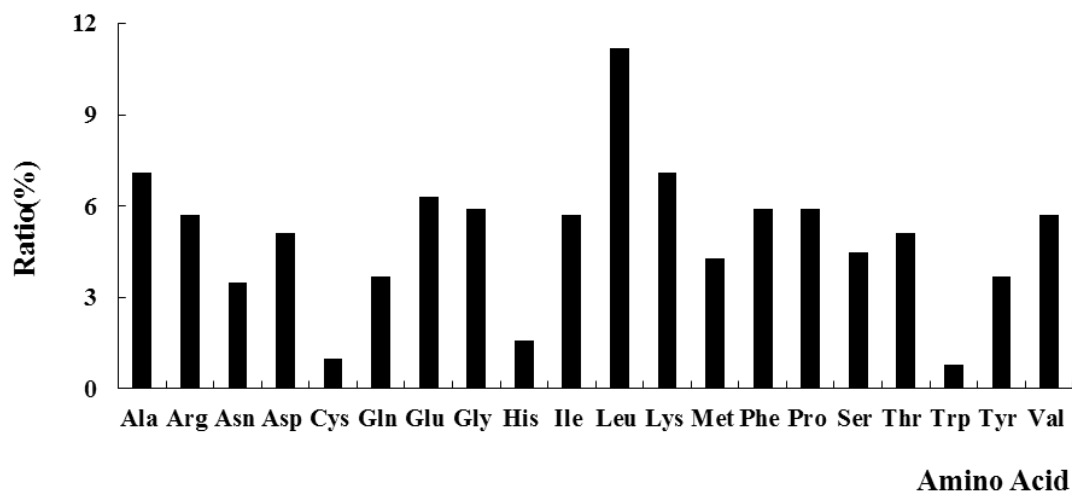


Fig. 2 Amino acid composition of BocCYP83B1

### 3.3 Subcellular Localization and Conserved Domain Analysis.

Subcellular localization of the *BocCYP83B1* gene was predicted by WoLF PSORT to be in the chloroplast. The analysis using Conserved Domain Database (CDD) demonstrated that the amino acid sequence of the BocCYP83B1 protein belonged the p450 superfamily.

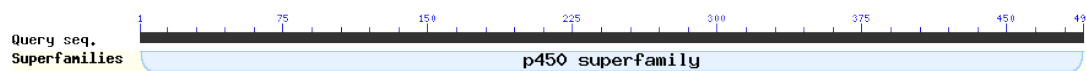


Fig. 3 Conserved domains analysis of BocCYP83B1

### 3.4 Homology and Phylogenetic Tree Analysis.

A phylogenetic tree was constructed to illustrate the relationship among the CYP83B1 proteins of cabbage and 18 other higher plant species (Fig. 4). A total of two major clusters were identified. Sequence alignment indicated that the BocCYP83B1 protein is more closely related to *B. rapa*.

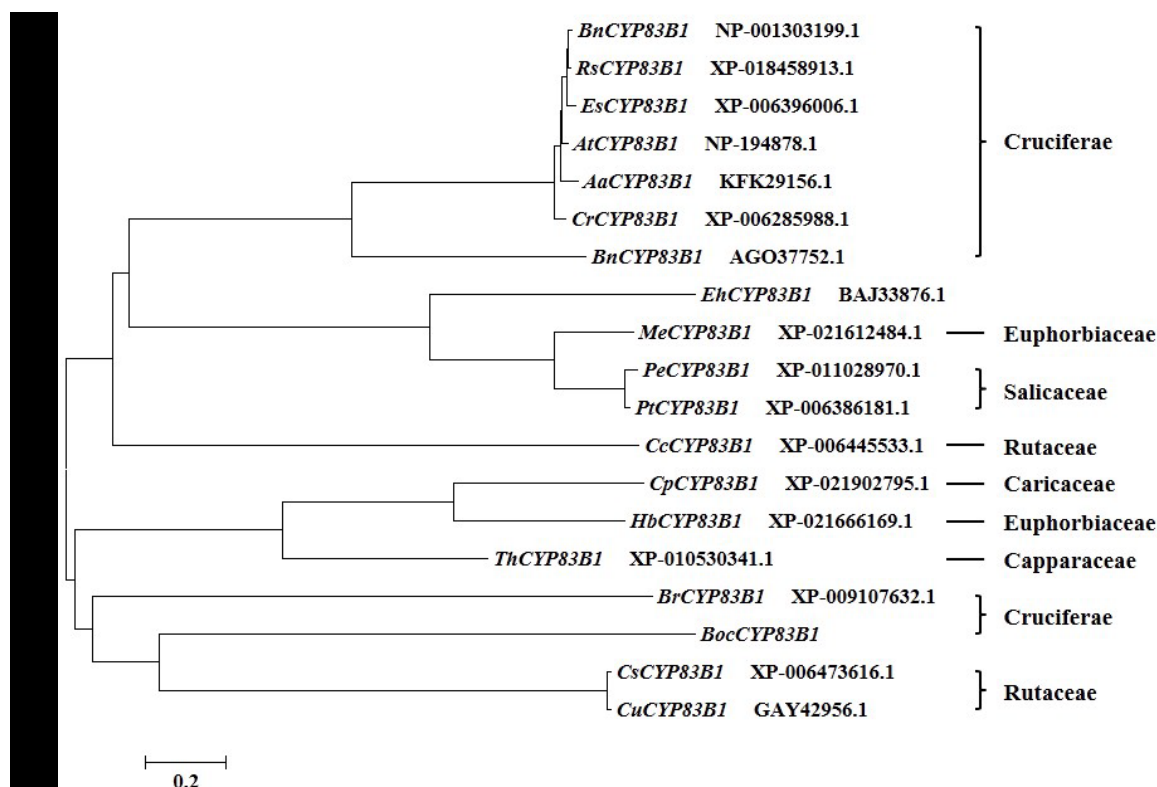


Fig. 4 Phylogenetic tree analysis of BocCYP83B1 and CYP83B1 proteins of other species

#### 4. Discussion

The present study analyzed the *BocCYP83B1* gene of cabbage. CYP83B1 enzyme is encoded by a single-copy gene in *Arabidopsis thaliana* [7]. While, the *CYP83B1* gene occurred as a single copy in cabbage, indicating that the enzyme may have undergone similar evolutionary patterns. Previous studies have shown that the CYP83B1 protein is relatively conserved in plants [7]. The findings of the present study show that CYP83B1 from cabbage is highly conserved in plants, similar to that observed in earlier reports. The findings of the present study may serve as a foundation for future studies on the functions of CYP83B1 in glucosinolate metabolism in cabbage.

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#### References

- [1] M. Wennberg, J. Ekvall, K. Olsson, and M. Nyman, Changes in carbohydrate and glucosinolate composition in white cabbage (*Brassica oleracea* var. *capitata*) during blanching and treatment with acetic acid, *Food Chem.* 95 (2006) 226-236.
- [2] S. Rokayya, C.J. Li, Y. Zhao, Y. Li, and C.H. Sun, Cabbage (*Brassica oleracea* L. var. *capitata*) phytochemicals with antioxidant and anti-inflammatory potential, *Asian Pac. J. Cancer Prev.* 14 (2014) 6657-6662.
- [3] B.G. Hansen, R.E. Kerwin, J.A. Ober, V.M. Lambrix, T. Mitchell-Olds, J. Gershenzon, B.A. Halkier, and D.J. Kliebenstein, A novel 2-oxoacid-dependent dioxygenase involved in the formation of the goiterogenic 2-hydroxybut-3-enyl glucosinolate and generalist insect resistance in *Arabidopsis*, *Plant Physiol.* 148 (2008) 2096-2108.
- [4] X. Yan, and S. Chen. Regulation of plant glucosinolate metabolism, *Planta* 226 (2007) 1343-1352.
- [5] E. Andréasson, L.B. Jørgensen, A.S. Höglund, L. Rask, and J. Meijer, Different myrosinase and idioblast distribution in *Arabidopsis* and *Brassica napus*, *Plant Physiol.* 127 (2001) 1750-1763.
- [6] N. Benkeblia, *Phytonutritional Improvement of Crops*, John Wiley & Sons, Inc., Hoboken, 2017, pp. 407-733.
- [7] S. Bak, F.E. Tax, K.A. Feldmann, D.W. Galbraith, and R. Feyereisen, CYP83B1, a cytochrome P450 at the metabolic branch point in auxin and indole glucosinolate biosynthesis in *Arabidopsis*, *Plant Cell* 13 (2001) 101-111.