Optimization of Fermentation Parameters for Laccase Production by a Novel Deuteromycete Fungus *Myrothecium verrucaria* NF-05 Using Response Surface Methodology

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Abstract—To obtain the maximum production of laccase from a novel deuteromycete fungus *Myrothecium verrucaria* NF-05, optimization of fermentation parameters was performed. Central composite design and response surface analysis revealed the optimum concentration of glucose, CuSO4 and gallic acid to be 26.47 g L⁻¹, 236.3 µM, and 138.4 µM, respectively. This optimization strategy led to the enhancement of laccase production from 12.00 to 19.94 U mL⁻¹, 1.66-fold increase.

Keywords—laccase; deuteromycete; medium; optimization; response-surface methodology

I. INTRODUCTION

Laccases (EC 1.10.3.2; benzenediol: oxygen oxidoreductase) are a family of multicopper oxidases that were first isolated from *Rhus venicifera*, the Japanese lacquer tree [1]. Laccases are able to catalyze direct oxidation of ortho- and para-diphenols, amino phenols, polyphenols, polyamines, and aryl diamines as well as some inorganic ions [2]. Due to its broad substrate specificity, laccase has great potential in varied environmental applications including pulp delignification, xenobiotics degradation and biopolymer bleaching [3-5]. Deuteromycetes fungi possess more advantages in regards to producing laccase, such as simple life cycle, short growth phase and facility to genetic reconstruction. Reports about laccase producing by deuteromycetes fungi mainly focus on lignin degradation [6, 7], inducement and purification [8-10] and decolorization of dye [11]. The significant factors which effected laccase production could be investigated by applying response surface methodology (RSM), which enables the study of interaction effects among different variables. Regression model could predict purpose response under untested sets of variables. RSM has been widely reviewed and proven to be efficacious for the production of industrial enzymes during fermentation of microorganisms [12-14].

Optimization of the fermentation process of laccase production by RSM has been reported in the case of different microorganisms [15, 16]. To the best of our best knowledge there are no reports studying the use of statistical design of experiments (DOE) in regards to the optimization of laccase from deuteromycetes. In this paper we report on the effectiveness of laccase production by a novel deuteromycete fungus *Myrothecium verrucaria* NF-05 and its optimization using DOE concepts.

II. MATERIALS AND METHODS

A. Microorganism

The microorganism used was deuteromycete fungus *Myrothecium verrucaria* NF-05, isolated from soil samples in Liangshui Nature Reserve, China and identified by the Department of Microbiology, College of Life Science, Northeast Forestry University. This strain was stored as spore suspension at -20°C in sterile 20% (v/v) glycerol solution.

B. Shake flake culture

All the cultures were grown in 250-mL Erlenmeyer flasks containing 60-mL fermentation liquid medium (potato 200 g L⁻¹, KH2PO4 1.0 g L⁻¹, MgSO4·7H2O 0.3 g L⁻¹). Concentrations of glucose, CuSO4 and gallic acid are 28 g L⁻¹, 230 µM L⁻¹ and 140µM L⁻¹ respectively. Each flask was inoculated with three 0.7-cm-diameter plugs taken from an agar plate (made up of basal potato-glucose medium, potato 200 g L⁻¹, glucose 20g L⁻¹, KH2PO4, 1.0 g L⁻¹, MgSO4·7H2O 0.3 g L⁻¹, agar 20 g L⁻¹), covered with fungal mycelium and grown for ten days. All cultures were incubated at 28°C.

C. Enzyme assay

Cell-free culture fluid, obtained after removal of mycelium by centrifugation (8000 rpm for 15 min), was used as the source of the enzyme. The activity of laccase was spectrophotometrically determined by applying 50 µL of culture filtrate which was diluted appropriately into 1 mL of 1 mM ABTS (2,2'-azino-bis-[3-ethyl benzothiazoline-6-sulphonic acid]) (SIGMA) in 0.2 M acetate buffer (pH 4.0) to get the absorbance in readable range. The reaction mixture (4 mL) was incubated in 30°C for 3 min based on Wu et al.’s method [17]. One unit of enzyme activity was defined as 1 µmol of ABTS oxidized per minute. To calculate enzyme activity the absorption coefficient ε420nm= 3.6×10⁴ M⁻¹ cm⁻¹ was used.

D. Experimental designs and data analysis

Laccase production can be written as a function

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(response surface) of coded values of variables with a significant influence on laccase production, and can be approximated by a second-order polynomial. CCD (Table 1) is a useful tool to acquire data to fit this polynomial (Y, U mL⁻¹). All the experiments were performed in triplicates according to a design matrix. The statistical software Design Expert (version 7.0.0, Stat-Ease, Minneapolis, USA) was used for experimental design and data analysis.

### III. RESULTS

The results of regression analysis are shown in Table 2. F-test for the analysis of variance done on the experimental data indicated that the model was highly significant, with F-value of 120.16 and P>F of 0.0001; the obtained model was adequate. The model had an $R^2$ value of 0.9914, indicating a good correlation between observed and predicted responses. Also, the model indicated that the predicted $R^2$ value of 0.9817 was in reasonable agreement with the adjusted $R^2$ value of 0.9155. The results demonstrated that glucose was found to have the most significant effect on laccase production by Myrothecium verrucaria NF-05, as indicated by a P>F-value of 0.0001.

**CuSO₄** and gallic acid had significant linear effects on the system. Interactions between glucose and CuSO₄ as well as CuSO₄ and gallic acid also had significant effects (P>F ≤ 0.05). The maximum point of the model can be obtained: 26.47 g L⁻¹ of glucose, 236.3 µM of CuSO₄ and 138.4 µM of gallic acid (coded level -0.51, 0.21 and -0.08, respectively). The model predicted a maximum response of 19.82 U/mL for this point. The results were confirmed by additional shake flask experiments using parameters representing this maximum point and a value of 19.94±1.16 U mL⁻¹ (N=6) was obtained. Good correlation between these two results verified the validity of the response model and the existence of an optimal point.

**FIGURE 1.** Contour plots of laccase production showing interactive effect of (a) CuSO₄ and Gallic acid (b) Glucose and CuSO₄ and (c) Glucose and Gallic acid

CuSO₄ together with its interactions with glucose and gallic acid had significant effects in this study (P>F-value of 0.0024, 0.0011, 0.0045). It is demonstrated that copper is an important inducer for laccase production which is in agreement with Gnanamani et al.’s report [21]. The yield of laccase was found to be higher with the coded level of CuSO₄ ranging from 0 to 0.5 and optimum concentration of 236.3 µM, which corresponds with the concentrations typically used in cultivation media (2-600 µM) for laccase production in wild-type or recombinant strains [22-24]. Several probable mechanisms have been proposed, i.e., regulating laccase expression at transcription level [18], acting in defense mechanism against oxidation pressure [25], formatting metallothioneins or cell-wall components [26]. The optimum concentration of copper varies remarkably among strains.

**TABLE 1 EXPERIMENTAL DESIGN AND RESULTS OF THE CENTRAL COMPOSITE DESIGN**

<table>
<thead>
<tr>
<th>Run</th>
<th>X₁</th>
<th>X₂</th>
<th>X₃</th>
<th>Laccase (U mL⁻¹) Observed</th>
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As an aromatic compound, gallic acid is a potential inducer of laccase [27, 28]. Gallic acid in a final concentration of 1 mM obviously advanced laccase activity by both *Streptomyces psammotiticus* [29] and white rot fungus WR-1 [28]. Nevertheless, higher levels of gallic acid seemed obstructive in this study. The optimum concentration was 138.4 µM around medium coded level when CuSO₄ was fixed at 0 to 0.5 of coded level. The deuteromycete fungus *Paecilomyces* sp. exhibits an evident decrease in laccase production under cooperative induction by 100 µM of gallic acid and the extra addition of 50 µM copper [9].

Laccase is the most extensively studied group of enzymes among oxidases and has attracted more and more interest for applications in environmental, food, bioelectronics and lignin degradation. To the best of our knowledge, *Myrothecium verrucaria* NF-05 is a newly isolated deuteromycete fungus strain without detailed study on fermentation process of laccase production. *Myrothecium verrucaria* NF-05 produced laccase within a short incubation period of 5-7 days and broad pH range of 6-8. These remarkable properties make this organism a better candidate for biotechnological applications especially in the areas where alkaline conditions are preferred. Based on present work, the characteristics of the laccase protein should be further studied; taking into consideration the synergistic inducing effects between CuSO₄ and gallic acid, and the adding time of inducers.

V. ACKNOWLEDGEMENT

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