Discrimination of Outer Membrane Proteins using Reformulated Support Vector Machine based on Neutrosophic Set

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Abstract

Neutrosophic logic is introduced in 1995 as a generalization of fuzzy logic. It includes a new component as neutralities. In this paper, we propose a novel neutrosophic set for SVM inputs and combine it with the reformulated SVM which treats samples differently according to the weighting function. The proposed classifier helps reducing the effects of outliers. We test it on discriminating outer membrane proteins (OMPs) from globular proteins and α-helical membrane proteins using amino acid composition and residue pair information. The experiment results show that the proposed method outperforms the traditional SVM in both classification accuracy and MCC.

Keywords: Neutrosophic set, reformulated support vector machine, outer membrane proteins (OMPs).

1. Introduction

Neutrosophic logic was introduced by Florentin Smarandache in 1995 as a generalization of fuzzy logic. It studies the neutrosophic logical values of the propositions. Each proposition is estimated to have three components: the percentage of truth in a subset $T$, the percentage of indeterminacy in a subset $I$, and the percentage of falsity in a subset $F$ [1]. Compared with all other logics, neutrosophic logic introduces a percentage of "indeterminacy" due to unexpected parameters hidden in some propositions. The main distinction between neutrosophic logic (NL) and fuzzy logic (FL) is that the sum of neutrosophic components in NL is not necessarily 1 as in FL but any number from 0 and 3 [2].

Support Vector Machine (SVM) developed by Vapnik and Cortes has superior features such as avoiding over-fitting and obtaining global optimal [3]. It has been applied to many problems in bioinformatics. Kim and Park [4] used it to predict protein relative solvent accessibility. Nguyen and Rajapakse [5] applied SVM to predict protein secondary structures. It has also been applied to protein domains identification (Vlahovicke et al., [6]), protein-protein binding sites prediction (Brandford and Westhead [7]), remote protein homology detection (Busuttil et al. [8]) and protein subcellular localization (Nair and Rost [9]).

Outer membrane proteins (OMPs) perform a variety of functions, such as selectively allowing the passage of molecules, mediating non-specific, passive transport of ions and small molecules [10]. Discriminating OMPs from globular proteins and α-helical membrane proteins is an important task both for dissecting OMPs...
from genomic sequences and for the successful prediction of their secondary and tertiary structures. Park et al. used SVM to discriminate OMPs based on amino acid composition and residue pair information in [10].

In this paper, we propose a novel neutrosophic set for the input samples of SVM. Combining the neutrosophic set with the reformulated SVM, we discriminate OMPs from globular proteins and α-helical membrane proteins. We use the same dataset in [10], which is composed of 208 OMPs, 673 globular proteins and 206 α-helical membrane proteins. The experimental results show that the proposed method outperforms the traditional SVM in both accuracy and MCC.

The rest of the paper is organized as follows. Section 2 introduces the reformulated support vector machine. Section 3 describes the proposed neutrosophic set for SVM input and how the neutrosophic set is integrated into the reformulated SVM in detail. The experiment results are listed in section 4 and conclusions are drawn in section 5.

2. Reformulated Support Vector Machine

SVM uses hypothesis space of linear functions in a high-dimensional feature space, and it is trained with a learning algorithm based on optimization theory [11].

Suppose we are given a training set \( S \) containing \( n \) labeled points \((x_i, y_i)\), where \( x_i \in \mathbb{R}^n \) and \( y_i \in \{-1, 1\}, i = 1, \ldots, n \). \( \Phi(x) \) denotes the mapping from \( \mathbb{R}^n \) to a feature space \( Z \). We want to find the hyperplane with maximum margin as:

\[
\mathbf{w} \cdot z + b = 0 \quad \text{(1)}
\]

such that for each point \((x_i, y_i)\), where \( z_\Phi(x_i) \),

\[
y_i (\mathbf{w} \cdot z_i + b) \geq 1, \quad i = 1, \ldots, n. \quad \text{(2)}
\]

When the data set is not linearly separable, the soft margin is allowed by introduction of \( n \) non-negative variables, denoted by \( \xi = (\xi_1, \xi_2, \ldots, \xi_n) \), such that the constraint for each sample in Eq. (2) is rewritten as:

\[
y_i (\mathbf{w} \cdot z_i + b) \geq 1 - \xi_i, \quad i = 1, \ldots, n. \quad \text{(3)}
\]

The optimal hyperplane problem is the solution to the problem

\[
\text{minimize} \quad \frac{1}{2} \mathbf{w} \cdot \mathbf{w} + C \sum_{i=1}^{k} \xi_i \quad \text{subject to} \quad y_i (\mathbf{w} \cdot z_i + b) \geq 1 - \xi_i, \quad i = 1, \ldots, n. \quad \text{(4)}
\]

where the first term in equation (4) measures the margin between support vectors and the second term measures the amount of misclassifications. \( C \) is a constant parameter that tunes the balance between the maximum margin and the minimum classification error.

Lin and Wang proposed fuzzy support vector machine in [12]. A membership \( s_i \) is assigned for each input sample \((x_i, y_i)\), where \( 0 < s_i < 1 \). Since the membership \( s_i \) is the attitude of the corresponding point \( x_i \) toward one class and the parameter \( \xi_i \) is a measure of error in the SVM, the term \( s_i \xi_i \) is a measure of error with different weighting. The optimal hyperplane problem is then regarded as the solution to

\[
\text{minimize} \quad \frac{1}{2} \mathbf{w} \cdot \mathbf{w} + C \sum_{i=1}^{k} s_i \xi_i \quad \text{subject to} \quad y_i (\mathbf{w} \cdot z_i + b) \geq 1 - \xi_i, \quad i = 1, \ldots, n. \quad \text{(5)}
\]

We use the similar idea in the reformulated SVM. The difference is that the membership \( s_i \) is substituted by weighting function \( g_i \), where \( g_i > 0 \). Different inputs contribute differently to the training procedure, and we use weighting function \( g_i \) to evaluate the degree of importance for each input. The value of \( g_i \) is a positive number and is unnecessary to be smaller.
than 1. Now the optimal hyperplane problem in the reformulated SVM is the solution to

\[
\begin{align*}
\text{minimize} & \quad \frac{1}{2} w \cdot w + C \sum_{i=1}^{k} g_i \xi_i \\
\text{subject to} & \quad y_i (w \cdot z_i + b) \geq 1 - \xi_i, \quad i = 1, \ldots, n.
\end{align*}
\]  

(8)

(9)

3. Integrating Neutrosophic Set with Reformulated SVM

3.1. Neutrosophic Set

Neutrosophic set is a generalization of the intuitionistic set, classical set, fuzzy set, paraconsistent set, dialetheist set, paradoxist set and tautological set [2].

In classical theory, there are only \(<A> and <\text{Non-A}>. The degree of neutralities \(<\text{Neut-A}> is introduced and added in neutrosophic theory. Generally a neutrosophic set is denoted as \(\langle T, I, F \rangle. An element \(x \in \langle t, i, f \rangle belongs to the set in the following way: it is \(t \) true in the set, \(i \) indeterminate in the set, and \(f \) false, where \(t, i, \) and \(f \) are real numbers taken from the sets \(T, I, \) and \(F, \) nor on their sum \(m=t+i+f. \) The major difference between neutrosophic set (NS) and fuzzy set (FS) is that there is no limit on the sum \(m \) in NS while in FS \(m \) must be equal to 1.

3.2. Proposed Neutrosophic Set for SVM input

Many research results have shown that the SVM is very sensitive to noises and outliers. Here we propose a neutrosophic set for the input samples of SVM based on the distances between the sample and the class centers. The reformulated SVM integrated with the proposed neutrosophic set can help solving the problems of noise and outliers.

Using the same notations in section 2, the neutrosophic set for input samples are denoted as a sequence of points:

\[
\begin{align*}
(x_j, y_j, t_j, i_j, f_j), \quad j = 1, \ldots, n.
\end{align*}
\]

Here for a sample \(x_j\) belongs to class \(y_j, \) it is \(t_j \) true, \(i_j \) indeterminate and \(f_j \) false. We define the center of positive samples \(C_+ \), the center of negative samples \(C_- \) and the center of all samples \(C_{all} \) as following:

\[
\begin{align*}
C_+ &= \sum_{k=1}^{n} x_k, \\
C_- &= \sum_{k=1}^{n} x_k, \\
C_{all} &= \sum_{k=1}^{n} x_k.
\end{align*}
\]

(10)

where \(n_+ \) is the number of positive samples and \(n_- \) is the number of negative samples.

We denote \(U\) as the whole input samples set, \(P\) as the positive samples subset and \(N\) as the negative samples subset. For positive samples where \(y_j = 1, \) the neutrosophic components are defined as:

\[
\begin{align*}
t_j &= 1 - \frac{||x_j - C_+||}{\text{max}_{x_k \in P} ||x_k - C_+||} \\
i_j &= 1 - \frac{||x_j - C_{all}||}{\text{max}_{x_k \in U} ||x_k - C_{all}||} \\
f_j &= 1 - \frac{||x_j - C_-||}{\text{max}_{x_k \in P} ||x_k - C_-||}
\end{align*}
\]

(11)

where \(||x||\) denotes the Euclidean distance of variable \(x.\)

For negative samples where \(y_j = -1, \) the neutrosophic components are defined as:

\[
\begin{align*}
t_j &= 1 - \frac{||x_j - C_-||}{\text{max}_{x_k \in N} ||x_k - C_-||} \\
i_j &= 1 - \frac{||x_j - C_{all}||}{\text{max}_{x_k \in U} ||x_k - C_{all}||} \\
f_j &= 1 - \frac{||x_j - C_+||}{\text{max}_{x_k \in N} ||x_k - C_+||}
\end{align*}
\]

(12)
With the above definition, every input sample is associated with a triple \(<t_j, i_j, f_j>\) as its neutrosophic components. The larger \(t_j\) it has, the more probability it belongs to the labeled class. The larger \(i_j\) it has, the more probability it is indeterminate. The larger \(f_j\) it has, the more probability it belongs to the opposite of the labeled class.

3.3. Integrating Neutrosophic Set with Reformulated SVM

In order to use the reformulated SVM, we should define a weighting function for input samples.

Following the steps in section 3.2, every sample has been associated with a triple \(<t_j, i_j, f_j>\) as its neutrosophic components. Larger \(t_j\) means the sample is nearer to the center of the labeled class and is less likely an outlier. So \(t_j\) should be emphasized in the weighting function. Larger \(i_j\) means the sample is harder to be discriminated between two classes. This factor should also be emphasized in the weighting function in order to classify the indeterminate samples more accurately. Larger \(f_j\) means the sample is more likely an outlier. This sample should be treated less importantly in the training procedure. Based on these analyses, we define the weighting function \(g_j\) as:

\[
g_j = t_j + i_j - f_j \tag{13}
\]

The reformulated SVM combined with the proposed weighting function treats samples differently in the training procedure and can help reducing the effects of outliers in the training samples.

4. Experimental Results

The same four measures in [10] are used to evaluate the classification performance. Sensitivity, specificity, overall accuracy and MCC are defined as

\[
sensitivity = \frac{TP}{TP + FN} \tag{14}
\]

\[
specificity = \frac{TN}{TN + FP} \tag{15}
\]

\[
overall accuracy = \frac{TP + TN}{TP + TN + FP + FN} \tag{16}
\]

\[
MCC = \frac{TP \times TN - FP \times FN}{\sqrt{(TP + FP)(TP + FN)(TN + FP)(TN + FN)}} \tag{17}
\]

where TP, FP, TN and FN refer to the number of true positives, false positives, true negatives and false negatives proteins, respectively.

We use the same dataset and repeat all the experiments with the same parameter \(\gamma\) as stated in [10]. There are three categories of experiments: discrimination of OMPs and globular proteins, discrimination of OMPs and \(\alpha\)-helical membrane proteins and discrimination of OMPs and non-OMPs. All of the experiments are results of 5-fold cross-validation test as in [10]. The results are listed in Tables 1, 2 and 3 respectively. The original results of [10] are listed in normal font while our results using FSVM are listed in Italic and Bold font with ‘F’ as postfix.

Table 1. Discrimination of OMPs and globular proteins.

<table>
<thead>
<tr>
<th>Input</th>
<th>Sen (%)</th>
<th>Spe (%)</th>
<th>Overall (%)</th>
<th>MCC</th>
</tr>
</thead>
<tbody>
<tr>
<td>20D</td>
<td>82.7</td>
<td>93.8</td>
<td>91.1</td>
<td>0.757</td>
</tr>
<tr>
<td>400D</td>
<td>82.9</td>
<td>95.4</td>
<td>92.5</td>
<td>0.793</td>
</tr>
<tr>
<td>400DF</td>
<td>85.1</td>
<td>97.7</td>
<td>95.2</td>
<td>0.863</td>
</tr>
<tr>
<td>420D</td>
<td>63.5</td>
<td>100</td>
<td>91.4</td>
<td>0.755</td>
</tr>
<tr>
<td>420DF</td>
<td>70.6</td>
<td>98.8</td>
<td>93.4</td>
<td>0.805</td>
</tr>
<tr>
<td>17D</td>
<td>87.5</td>
<td>94.1</td>
<td>92.5</td>
<td>0.798</td>
</tr>
<tr>
<td>17DF</td>
<td>91.6</td>
<td>95.3</td>
<td>92.9</td>
<td>0.829</td>
</tr>
</tbody>
</table>
Table 2. Discrimination of OMPs and α-helical membrane proteins.

<table>
<thead>
<tr>
<th>Input</th>
<th>Sen (%)</th>
<th>Spe (%)</th>
<th>Overall (%)</th>
<th>MCC</th>
</tr>
</thead>
<tbody>
<tr>
<td>20D</td>
<td>98.6</td>
<td>91.3</td>
<td>94.9</td>
<td>0.901</td>
</tr>
<tr>
<td>20DF</td>
<td>98.8</td>
<td>92.9</td>
<td>95.7</td>
<td>0.913</td>
</tr>
<tr>
<td>400D</td>
<td>99.0</td>
<td>90.3</td>
<td>94.7</td>
<td>0.897</td>
</tr>
<tr>
<td>400DF</td>
<td>98.7</td>
<td>92.6</td>
<td>95.6</td>
<td>0.918</td>
</tr>
<tr>
<td>420D</td>
<td>95.7</td>
<td>89.8</td>
<td>92.8</td>
<td>0.856</td>
</tr>
<tr>
<td>420DF</td>
<td>95.1</td>
<td>92.9</td>
<td>93.8</td>
<td>0.881</td>
</tr>
<tr>
<td>15D</td>
<td>99.0</td>
<td>92.7</td>
<td>95.9</td>
<td>0.920</td>
</tr>
<tr>
<td>15DF</td>
<td>99.1</td>
<td>93.4</td>
<td>96.6</td>
<td>0.928</td>
</tr>
</tbody>
</table>

Table 3. Discrimination of OMPs and non-OMPs.

<table>
<thead>
<tr>
<th>Input</th>
<th>Sen (%)</th>
<th>Spe (%)</th>
<th>Overall (%)</th>
<th>MCC</th>
</tr>
</thead>
<tbody>
<tr>
<td>20D</td>
<td>87.5</td>
<td>92.6</td>
<td>91.6</td>
<td>0.752</td>
</tr>
<tr>
<td>20DF</td>
<td>87.8</td>
<td>93.7</td>
<td>93.1</td>
<td>0.792</td>
</tr>
<tr>
<td>400D</td>
<td>86.5</td>
<td>96.4</td>
<td>94.5</td>
<td>0.823</td>
</tr>
<tr>
<td>400DF</td>
<td>86.8</td>
<td>97.2</td>
<td>94.9</td>
<td>0.848</td>
</tr>
<tr>
<td>18D</td>
<td>89.9</td>
<td>92.5</td>
<td>92.0</td>
<td>0.767</td>
</tr>
<tr>
<td>18DF</td>
<td>90.3</td>
<td>94.9</td>
<td>94.1</td>
<td>0.821</td>
</tr>
<tr>
<td>28D</td>
<td>90.9</td>
<td>94.7</td>
<td>93.9</td>
<td>0.816</td>
</tr>
<tr>
<td>28DF</td>
<td>89.7</td>
<td>96.3</td>
<td>94.8</td>
<td>0.831</td>
</tr>
</tbody>
</table>

Due to the page settings, some notations are abbreviated in the tables. Sensitivity is truncated as sen while specificity is denoted by spe. The results show that the proposed method outperforms the traditional SVM in both overall classification accuracy and MCC. The increase of overall accuracy is 1%-2% and the MCC is increased by 3%-4% in most cases. The improvement is significant and adequately validates the correctness and effectiveness of the proposed method.

5. Conclusion

SVM is sensitive to outliers and noises in the input samples. In order to eliminate the effects of outliers, we integrate reformulated SVM with neutrosophic set derived from input samples. The reformulated SVM treats samples differently according to the weighting function in the training procedure. The weighting function is based on the neutrosophic set. We apply the proposed method to the discrimination of outer membrane proteins and compare the results with that of the traditional SVM. The experimental results have shown that the proposed classifier achieves higher accuracy and MCC than the traditional SVM method does.

6. References


