Prediction of Enantioseparation of Econazole on the Cyclodextrin Derivatives as Chiral Selectors by Molecular Docking Approach

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ABSTRACT

Molecular docking approach has been successfully developed for prediction of enantioseparation of econazole on the cyclodextrin derivatives namely sulfated-β-cyclodextrin (S-β-CD) and hydroxypropyl-γ-cyclodextrin (HP-γ-CD) as chiral selectors. Molecular docking was performed using AutoDock Vina software and the root mean square deviation (RMSD) was calculated using PyMol software. Molecular docking shows that R-Econazole forms more stable interaction with all cyclodextrin derivatives than S-Econazole forms, suggesting that S-econazole will be eluted earlier than R-econazole. In addition, the stability level of cyclodextrin derivatives as chiral selectors was S-β-CD > HP-γ-CD.

Keywords: Cyclodextrin, Enantioseparation, Econazole, Molecular docking.

1. INTRODUCTION

Econazole is an antifungal agent well-known for the potential treatment of several fungal infections in the skin of both humans and animals. This compound has one chiral center, thus leading to two enantiomers. The activity and toxicity of chiral azole drugs might be influenced by stereoisomerism. Based on several reports, there is only one enantiomer that gives good effects [1]. Therefore, the enantiomeric separation of chiral compounds is important to do. One of the most possible methods to separate enantiomers of chiral compounds is high-performance liquid chromatography (HPLC) and capillary electrophoresis (CE) [2-4].

Chiral separation can be achieved using chiral selectors which clearly distinguish the two enantiomers. Chiral selector changes one of the two enantiomers at different rates into a new compound (kinetics enantioselective) or established a labile molecule at different stability (thermodynamics enantioselective). One of the most widely used chiral selectors is cyclodextrin due to its excellent chiral recognition abilities [5]. Cyclodextrin (CD) is a molecule that has a cyclic torus shape that has an outer surface hydrophilic and a lipophilic middle cavity that can accommodate various lipophilic ligands. Cyclodextrin is a macromolecule consisting of glucopyranose subunits obtained by enzymatic degradation of starch. Cyclodextrin has a conical shape with a lipophilic center cavity and a hydrophilic surface [6-7]. However, the HPLC chiral separation can only separate a limited chiral range. Therefore, a preliminary schematic was needed to analyze the three-dimensional structure of compounds to predict the characteristics and functions of these compounds and analyze the actual interactions at the molecular level [8].

Molecular docking is a computational method that has been used to predict the molecular interactions of ligands and receptors. The molecular docking prediction increases accuracy and precision in chiral recognition and also reduces research time since it can be predicted quickly [9]. The molecular docking method can save time, effort, and the use of solvents or chemicals that pollute the environment [10]. This method was performed by placing the ligands systematically on the active site of receptors. The molecular docking method aims to achieve the optimal complex conformation and to predict the interactions of drugs/ligands and receptors, therefore, the best geometry of ligands and receptors.
receptors complex was obtained. In addition, a more effective ligand can be predicted by calculating the energy interactions of the different ligands [11]. Based on this report, this study aims to analyze the structure of the econazole complex, calculate the binding energy, and determine the most suitable chiral selector to predict the separation of the enantiomers of econazole. Chiral selectors commonly used are cyclodextrin and its derivatives. Cyclodextrin is an oligosaccharide composed of six (α-CD), seven (β-CD), and eight (γ-CD) glucose units through α-1,4 glycoside bonds [12-15]. Cyclodextrin can form an inclusion complex with drugs by inserting drug molecules into the central cavity of cyclodextrin. The inclusion complex formed can improve the solubility, dissolution, stability, and bioavailability of the guest molecule [16]. Cyclodextrin shows good enantioselectivity for a wide range of analytes, is transparent to UV light and has good water solubility. In addition, cyclodextrins are available in various ranges, generally, give fast kinetics for the complex formation and breakdown enantiomers, and are relatively cheap [15]. In this study, econazole acts as ligands, and cyclodextrin acts as receptors. Cyclodextrins used for this study are sulfated-β-cyclodextrin (S-β-CD) and hydroxypropyl-γ-cyclodextrin (HP-γ-CD) [17].

2. MATERIALS AND APPARATUS

The crystal structure of S-β-CD and HP-γ-CD were obtained from PubChem (CID: 12049144 and 2733545) in SDF format. The crystal structure of R-Econazole and S-Econazole were obtained from PubChem (CID: 6604378 and 12773795).

A computer with an Operating System (OS) Windows 10 Enterprise 64-bit with processor Intel® Core™ i3-4005U CPU @ 1.70 GHz (4 CPUs), 1.7 GHz and external hard disk 500 GB, RAM 4 GB. The CDs structures were converted into PDB file format using Avogadro software. The structure of Econazole was optimized using the PM3 method implemented in HyperChem software. The AutoDock Vina software was used as a molecular docking tool. Docking results were calculated RMSD using PyMOL software.

2.1. Geometry Optimization

The structure of econazole was geometrically optimized using HyperChem software. The calculation used is semiempirical with PM3 as the method. The structure of econazole and its enantiomers are shown in Figure 1.

![Figure 1 Structure of R-Econazole and S-Econazole](Image)

2.2. Molecular Docking

Molecular docking was carried out using an automated docking program, AutoDock Vina. Water molecules on the receptor structures are removed to avoid interference during the molecular docking process. Polar hydrogen atoms are added to the structure as only polar atoms will interact with ligands while non-polar hydrogen atoms will be hidden to increase molecular docking calculation speed. The grid box was determined according to the coordinates of the active site of receptors. This study was conducted using a blind docking process as the grid box parameters that are not yet known. The large grid box was arranged so that ligands can rotate freely to find the most stable site on the receptors.

Exhaustiveness was a parameter in Autodock Vina that can control some of its comprehensive predictions. The increase of the exhaustiveness value will be slow down the docking process. However, the higher the exhaustiveness, the higher the probability of a good result of docking. The default value of Autodock Vina exhaustiveness is 8 [6,19]. In this study, exhaustiveness of 264 was used to obtain a more consistent docking result. The S-β-CD structure was docked with coordinate value of X = -0.437, Y = 9.514, Z = -8.760, Grid Box size of 20 x 20 x 20 and Grid Spacing 1.000. The HP-γ-CD structure was docked with coordinate value of X = -20.980, Y = 2.342, Z = 67.004, Grid Box size of 18 x 18 x 18 and Grid Spacing 1.000. The docking results should have a Root Mean Square Deviation (RMSD) of less than 3 Å [23].

3. RESULTS

3.1. Geometry Optimization

Geometry optimization can determine the location of atoms in a stable molecular conformation with the lowest energy state to obtain a molecular geometry that is a representation of the molecular structure adopted by compounds in nature.

The first step is ligands and receptors preparation using Avogadro software to change the file format into PDB. Econazole is a ligand that was optimized using HyperChem software by the semiempirical PM3 method, therefore, the most stable structure that has the lowest binding energy was obtained. Econazole enantiomers structures before and after optimization were shown in Figure 1 and Figure 2.

![Figure 2 The structure of R-Econazole before and after optimization](Image)

![Figure 3 The structure of S-Econazole before and after optimization](Image)
3.2. Molecular Docking

Molecular Docking is a computational simulation used to predict between a drug/ligand and a receptor/protein by attaching a small molecule (ligand) to the active site of the receptor [12]. This method was performed by placing the ligands systematically on the active site of receptors. The molecular docking method aims to achieve the optimal complex conformation and to predict the interactions of drugs/ligands and receptors, therefore, the best geometry of ligands and receptors complex was obtained. In addition, a more effective ligand can be predicted by calculating the energy interactions of the different ligands [16].

The running process was performed using Autodock Vina software. Water molecules on the receptor structures are removed to avoid interference during the molecular docking process. Polar hydrogen atoms are added to the structure as only polar atoms will interact with ligands while non-polar hydrogen atoms will be hidden to increase molecular docking calculation speed. The grid box was determined according to the coordinates of the active site of receptors. This study was conducted using a blind docking process as the grid box parameters are not yet known. The large grid box was arranged so that ligands can rotate freely to find the most stable site on the receptors.

Exhaustiveness was a parameter in Autodock Vina that can control some of its comprehensive predictions. The increase of the exhaustiveness value will be slow down the docking process. However, the higher the exhaustiveness, the higher the probability of a good result of docking. The default value of Autodock Vina exhaustiveness is 8 [20,23]. In this study, exhaustiveness of 264 was used to obtain a more consistent docking result. The RMSD values of inclusion complexes obtained are shown in Table 1. All of the RMSD values obtained are below 3 Å, this indicates that the ligand structure before and after docking is almost accurate.

### Table 1. RMSD value of the docking results.

<table>
<thead>
<tr>
<th>Inclusion complex</th>
<th>RMSD value (Å)</th>
</tr>
</thead>
<tbody>
<tr>
<td>S-β-CD/R-Econazole</td>
<td>1.509</td>
</tr>
<tr>
<td>S-β-CD/S-Econazole</td>
<td>1.667</td>
</tr>
<tr>
<td>HP-γ-CD/R-Econazole</td>
<td>1.642</td>
</tr>
<tr>
<td>HP-γ-CD/S-Econazole</td>
<td>2.371</td>
</tr>
</tbody>
</table>

The results of molecular docking are shown the stability of the inclusion complex based on its binding energy (ΔG). Figure 4 shows the structure of S-β-CD/R-Econazole and S-β-CD/S-Econazole complexes. The complex which the econazole is not included in the S-β-CD has ΔG value of S-β-CD/R-Econazole in the range -5.2 to -4.4 Kcal/mol and S-β-CD/S-Econazole in the range -4.7 to -4.2 Kcal/mol. The S-β-CD/R-Econazole complex has a lower binding energy value (-5.2 Kcal/mol) compared to S-β-CD/S-Econazole (-4.7 Kcal/mol). Econazole that does not include the S-β-CD may be caused by the size of the S-β-CD cavity, therefore, the complexes between econazole and S-β-CD could be trapped. The ΔG value of S-β-CD/R-Econazole and S-β-CD/S-Econazole docked using Autodock Vina software are shown in Table 2 and Figure 4. The negative value of ΔG indicates that the inclusion complex formed is stable [24].

### Table 2. The ΔG value of S-β-CD/R-Econazole and S-β-CD/S-Econazole.

| Inclusion complex       | ΔG (Kcal/mol) | |ΔΔG| (Kcal/mol) |
|-------------------------|---------------|----------------|---------------|
| S-β-CD/R-Econazole      | -5.2          | 0.5            |
| S-β-CD/S-Econazole      | -4.7          |                |

Figure 4 Structure of S-β-CD/R-Econazole and S-β-CD/S-Econazole.

The more negative the ΔG value, the higher affinity between ligands and receptors, therefore, the inclusion complex obtained was more stable. The enantiomer with the highest affinity will spend the most time in the mobile phase while the enantiomer with the least affinity will be eluted first from the column. The |ΔΔG| value is the difference ΔG value of R-Econazole and S-Econazole that describes the enantiomer separation ability to econazole compounds using chiral selector cyclodextrin. The value of |ΔΔG| obtained is 0.5 Kcal/mol. The higher |ΔΔG| value of |ΔΔG|, the better separation of chiral compounds. The value of |ΔΔG| indicates that cyclodextrin can be used as a chiral selector to separate the enantiomer of econazole compounds. In this study, the inclusion complex of R-Econazole was lower ΔG values compared to S-Econazole. It causes S-Econazole will be eluted first from the cyclodextrin chiral selector because the stability of S-Econazole is weaker than R-Econazole [25-26].

Following the same approach, the ΔG value of the inclusion complex was obtained from the docking process using Autodock Vina. The ΔG value of HP-γ-CD/R-Econazole is in the range -4.2 to -3.7 Kcal/mol and HP-γ-CD/S-Econazole is in the range -4.1 to -3.6 Kcal/mol. HP-γ-CD/R-Econazole complex has a lower binding energy value (-4.2 Kcal/mol) compared to HP-γ-CD/S-Econazole (-4.1 Kcal/mol). This complex has |ΔΔG| value in the range 0.1 Kcal/mol. In addition, HP-γ-CD/R-Econazole was a lower ΔG value compared to S-Econazole, therefore, S-Econazole will be eluted first from HP-γ-CD chiral selector.

### Table 3. The ΔG value of HP-γ-CD/R-Econazole and HP-γ-CD/S-Econazole.

| Inclusion complex       | ΔG (Kcal/mol) | |ΔΔG| (Kcal/mol) |
|-------------------------|---------------|----------------|---------------|
| S-β-CD/R-Econazole      | 1.509         |                | 0.1           |
| S-β-CD/S-Econazole      | 1.667         |                |               |
4. DISCUSSION

The molecular docking of econazole and cyclodextrin derivatives showed negative energy and a small number of RMSD. From the docking test, the RMSD value was less than 3 Å, so the prediction was quite accurate. The complexing ability of cyclodextrin was modified by the presence of hydroxypropyl (HP) substituent, therefore, the interaction and complexion energies for S-β-CD are higher than other cyclodextrin derivatives [27-28]. Based on the research that has been done, the use of S-β-CD as a chiral selector for chiral separation of econazole obtained the lowest AG value with high affinity and stable complex. It indicates that S-β-CD performed better than HP-γ-CD as a chiral selector for chiral separation of econazole compounds [29-31].

A high performance liquid chromatography (HPLC) method using cyclodextrin-based Astec Cyclobond as a chiral column has been successfully developed for chiral separation of econazole with Rs = 2.29 and analysis time within 9 min. Condition mobile phase composition of acetonitrile: water (0.2% HCOOH) 20:80 (v/v), and UV detection at 220 nm. The calibration curve of econazole was linear with \( r^2 = 0.9992 \). LOD and LOQ of econazole were 3.31 and 11.03 mg/L, respectively.

5. CONCLUSION

Cyclodextrin derivatives were used to predict chiral separation of econazole compounds, specifically sulfated-β-cyclodextrin (S-β-CD) and hydroxypropyl-γ-cyclodextrin (HP-γ-CD). The results of molecular docking using Autodock Vina shows that R-Econazole has a more stable interaction with all of the cyclodextrin derivatives compared to S-Econazole based on its lower binding energies. These results indicate that S-Econazole will be eluted first followed by R-Econazole. In addition, the stability level of cyclodextrin derivatives as a chiral selector to predicting the separation of econazole is S-β-CD > HP-γ-CD.

AUTHORS’ CONTRIBUTIONS

ALL AUTHORS made substantial contributions to CONCEPT and DESIGN, ACQUISITION OF DATA, or ANALYSIS and INTERPRETATION OF DATA; took part in DRAFTING the article or REVISING it critically for important intellectual content; agreed to submit to the current journal; gave final approval of the version to be published; and agree to be accountable for all aspects of the work. All the authors are eligible to be an author as per the international committee of medical journal editors (ICMJE) requirements/guidelines.

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